# The immune response of different farm animals vaccinated with bivalent FMD vaccine

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Cellular and humoral immune responses to bivalent FMD vaccine were estimated in vaccinated cattle, buffaloes, sheep, goats and camels. Five animals of each species were vaccinated using a dose of 1 ml for sheep and goats and 2 ml for cattle, buffaloes and camels inoculated subcutaneously. It was found that both of the immune parameters were generally increased gradually to reach the highest value of cellular immunity by the 28th day, while that of neutralizing antibodies was reached by the 6th week to the 10th week post vaccination. It was noticed that vaccination of cattle and sheep resulted in higher and longer duration of immune response than those obtained in vaccinated goat and buffaloes. While in vaccinated camels there was no evidence of antibodies against FMD. On the other hand, there were no significant differences between the levels of cellular immunity in different vaccinated animal species.

Key words: FMD-vaccine,

### INTRODUCTION

Rotavirus is a nonenveloped segmented (11 segments), double-stranded RNA (dsRNA) virus belonging to family Reoviridae. Rotaviruses are the single most important etiological agent of diarrheal disease in infants and young animals throughout the world (Estes, 2001). The rotavirus

virion consists of a three concentric protein shells, or layers. innermost layer, core, or composed of the VP2 protein, the middle layer is composed of the VP6 protein, and the outer layer is composed of the major surface glycoprotein, VP7 (glycoprotein, G) and haemagglutinin spike, VP4 (Protease sensitive, P); enclose neutralization antigens (Estes and Cohen, 1989).

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infection of cattle, pigs, sheep, goats, buffaloes and artiodactyls wildlife species. It is characterized by fever; vesicles in mouth and on the muzzle, teats and feet; and death in young animals (OIE annual status, 2006). All species of deer and antelope, elephant and giraffe are susceptible to FMD but camels are resistant to natural infection. Smaller camelids such as alpacas and llamas, although susceptible, are probably of no epidemiological significance. Mice and guinea pigs can be infected experimentally (Armstrong et al., 1998 and Moussa et al., 1998).

Also dromedaries can contract the disease after experimental infection or through close contact with FMD diseased livestock but do not present a risk in transmitting FMD to susceptible animals (Warnery et al., 2004 and Shawky et al., 2006).

It was stated that serotypes O and A of FMD virus has been isolated from sheep and goats (Kitching et al., 2002 and Assem, 2006). In addition, serological surveys revealed an evidence of occurrence of the disease in buffaloes (Starver et al., 1970; Abu Baker, 1996 and Abeer et al., 2002). The disease is considered enzootic in Egypt and many outbreaks have recurrently

occurred involving most governorates (Moussa et al., 1974; Daoud et al., 1988; El-Nakashly et al., 1996 and Farrag et al., 2004 and 2005). The causative serotype in such outbreaks was mainly type O but the last outbreak was found to be due the type A of FMD virus (Abdel-Rahman et al., 2006).

A new locally inactivated bivalent vaccine was generated containing the two types of the virus. Such vaccine was found to be safe and potent and helps to overcome the challenge and natural infection of animals with the virulent viral strains. Both cellular and humoral immune responses of animals usually share crucial role in the protection against FMD where the first one appears mainly more rapid than the second one but last Investigation of the shorter. immunogenicity of the bivalent FMD vaccine showed that it is a potent and offers a protective immunity in vaccinated animals (Abd El-Rahman et al., 2007).

The present work was designed to investigate and evaluate the cellular and humoral immune response to the bivalent FMD vaccine in cattle, buffaloes, sheep, goats and camels.

# MATERIAL AND METHODS

### 1- Animals:

Five male Balady sheep and goats of about 6 months of age, were purchased from El-Wadi El-Gedid, weighing about 50-60 Kgm. Male native breed cattle and buffaloe calves of about 6-9 months of age and male camel of 9 months age, were screened and found to be free from antibodies against FMD using serum neutralization test.

These animals were vaccinated with the local inactivated bivalent FMD vaccine in a dose of 1 ml for sheep and goats, 2 ml for cattle, buffaloes and camel, inoculated subcutaneously. In addition 2 animals of each species were kept without vaccination as control.

Serum samples were collected from all animals every week post vaccination till the 4<sup>th</sup> week, then every 2 weeks till the 20<sup>th</sup> week, to estimate the humoral immunity. In addition to heparinized blood samples which were collected from all animals on the 3<sup>rd</sup>, 7<sup>th</sup>, 14<sup>th</sup>, 21<sup>st</sup>, 28<sup>th</sup> and 35<sup>th</sup> days post vaccination to evaluate the cellular immunity.

2- Inactivated bivalent foot and mouth disease vaccine:

It was supplied by the Department of foot and mouth disease vaccine research, Veterinary Serum and Vaccine Research Institute, Abbassia, Cairo. It was used for vaccination of experimental animals.

## 3- FMD virus strains:

BHK cell culture adapted FMD virus serotypes O and A were supplied by the same department and used in SNT.

# 4- Serum Neutralization test (SNT):

The obtained samples from vaccinated animals were inactivated at 56°C for 30 minutes then subjected to SNT according to Ferreira (1976).

# 5- Lymphocyte proliferation test (L.P.T):

It was carried out as described by Lucy (1984) and modified by Abeer et al. (2002).

# 5.1. Phytohaemagglutinin (CPH):

It is none specific mitogen supplied by Biochrom KG, Leo Renstr, 2-6-D-1224, Berlin, Germany.

- 5.2. Roswell Park Memorial Institute, 1640 medium (RPMI-1640).
- 5.3. Ficol solution.

5.4. (4, 5 dimethylthiazol-2-y1,2, 5-diphenyltetra-zolium bromide (MTT).
5.5.Sodium dodecyle sulphate (SDS):

It were supplied by Sigma Pharmaceutical company.

### Results and Discussion

The obtained results of the lymphocyte blastogenesis assay measuring the cell mediated immune response of vaccinated animals showed that the delta optical density (ΔOD) correspond phytohaemagglutinin specific mitogen) were increased gradually from the 3rd week post vaccination (WPV) to reach its highest value by the 28th day then began to decline by the 35th day. Using FMDV as specific mitogen the  $\triangle OD$  values were higher than corresponding those phytohaemagglutinin. This behavior of such immune response was nearly the same among all vaccinated animals but it seems to be of no value in camel when FMDV was used as specific while use mitogen of phytohaemagglutinin as nonspecific mitogen reflected in stimulating of cellular immunity.

These findings appear to be in agreement with those of Halima et al. (1999), Abeer et al. (2002) and Abd El-Rahman et al. (2007) who showed that cellular immunity plays an important role in the immune response to the inactivated foot and mouth disease vaccine Also the observed findings in vaccinated camels that produce antibodies were supported by those of Shawky et al. (2006) who reported that camel can contract FMD after experimental infection but do not produce serum antibodies as in the case of vaccination.

On the other side. serum neutralization test revealed that **FMD** neutralizing specific antibodies were detectable from the 1<sup>st</sup> week post vaccination and reached the highest level by the 10<sup>th</sup> WPV in vaccinated cattle and sheep  $(2.7 \log_{10} - 2.25 \log_{10})$  and by the 8th WPV in buffaloes and goats  $(1.95 \log_{10} - 1.95 \log_{10})$  but in vaccinated camels there were no serum neutralizing developing antibodies. The mean serum neutralizing antibody titers in vaccinated cattle and sheep was still protective till the 16<sup>th</sup> WPV  $(1.5 \log_{10} - 1.2 \log_{10})$  while it was still protective till the 14<sup>th</sup> WPV in goats  $(1.2 \log_{10})$ , while

vaccinated buffaloes the mean serum neutralizing antibody titer still protective till the 12<sup>th</sup> WPV (1.2 log<sub>10</sub>).

These findings come parallel to those recorded by Barteling and Vreeswijk (1991) and Abd El-Rahman et al. (2006) who found that the highest mean titer of FMD antibodies in vaccinated cattle was obtained at 8th - 10th WPV with a duration of immunity up to 16 WPV. Also Halima et al. (1999) and Abeer et al. (2002) reported that in FMD vaccinated sheep the mean serum neutralizing antibody titer reached the highest level within 10th WPV and still protective up to 16th WPV. Assem (2006) and Fathia (2003) mentioned that the highest mean FMD serum neutralizing antibody

titer was reached at 8<sup>th</sup> WPV in foot and mouth disease vaccinated goats and was still protective till the 12<sup>th</sup> to 14<sup>th</sup> WPV.

In addition Abu Bakr (1996) and Abeer et al. (2002) found that the duration of humeral immunity induced by foot and mouth disease vaccine in vaccinated buffaloes was 12 WPV, then became within unprotective titer. Shawky et al. (2006) reported that experimentally infected and vaccinated camels have no ability to induce antibodies against FMD virus and FMD vaccine.

From the obtained results, it could be concluded that there is a variation in both humeral and cellular immune response to the FMD inactivated bivalent vaccine among various species.

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Table (1) Mean FMD type A serum neutralizing antibody titers (expressed as log10) in different animal species vaccinated with inactivated bivalent FMD vaccine

Animal	7.02		Mea	n FMD t	ype A an	tibody ti	Mean FMD type A antibody titers on week intervals post vaccination	eek inter	vals post	vaccinat	ion		
species	Prevac.	*-	2	3	4	9	8	10 12	12	14	9	18	20
Cattle	0.3	9.0	1.05	1.35	1.65	1.95	1.05 1.35 1.65 1.95 2.4 2.7	2.7	2.25	8.1	1.5	1.05 0.75	0.75
Buffaloe	0.15	0.45	6.0	1.2	1.35	1.65	1.65 1.95	1.5	1.2	6.0	0.9 0.75	0.45	0.3
Sheep	0.3	9.0	6.0	1.2	1.5		1.65 1.95 2.25	2.25	1.95	1.95 1.65 1.2	1.2	6.0	9.0
Goat	0.3	9.0	6.0	1.2	1.35	1.65	1.35 1.65 1.95	1.8	1.5	1.5 1.2	1.05 0.75	0.75	0.3
Camel	0.15	0.3	0.3	0.15	0.15 0.15	0.0	0.3	0.15	0.3	0.3	0.0	0.15	0.3

\* = weeks post vaccination

# Abeer, E.Mansour and Hegazi, A.Z.

Table (2) Mean FMD type O serum neutralizing antibody titers (expressed as log<sub>10</sub>) in different animal species vaccinated with the inactivated bivalent FMD vaccine

Animal		Me	Mean FMD type A antibody titers on week intervals post vacci	type A	antiboo	dy titers	on wee	k interv	als pos	t vacci	nation	-	
species	Prevac.	1*	2	ω	4	6	8	10	12	14	16	18	20
Cattle	0.15	0.45 0.9		1.2	1.5	1.8	2.1	2.4 2.1	2.1	1.8	1.5	1.5	0.6
Buffaloe	0.15	0.3	0.6	1.2	1.2 1.35 1.5		1.8	1.5	1.2 0.75	0.75	0.65	0.65 0.45	0.3
Sheep	0.15	0.45	0.75 1.2 1.35 1.65 1.95 2.1	1.2	1.35	1.65	1.95	2.1	1.8	1.5	1.2	0.9	0.45
Goat	0.3	0.6	0.6	1.2	0.6 1.2 1.35 1.65 1.8 1.65 1.5 1.2	1.65	1.8	1.65	1.5	1.2	0.9	0.6	0.3
Camel	0.15	0.15	0.3	0.3	0.3 0.3 0.15 0.0 0.0 0.3 0.15	0.15	0.0	0.0	0.3	0.15	0.3	0.3	0.0

 <sup>=</sup> weeks post vaccination

The immune response of different farm animals

Table (3) Lymphocyte proliferation test in different animal species vaccinated with inactivated

bivalent FMD vaccine	of lymphocyte proliferation / days post vaccination	3 DPV 7 DPV 14 DPV	0.200 0.235 0.312	0.241 0.269 0.341	0.194 0.228 0.305	0.245 0.277 0.334	0.271 0.242 0.326	0.261 0.281 0.415	0.205 0.238 0.318	0.253 0.258 0.420	0.201 0.225 0.275	0.105 0.109 0.103
	1/days post vace	V 21 DPV	0.380	1 0.405	5 0.365	4 0.397	6 0.367	5 0.412	8 0.371	0 0.411	5 0.319	3 0.105
	cination	28 DPV	0.425	0.456	0.400	0.439	0.431	0.460	0.409	0.448	0.400	0.100
		35 DPV	0.287	0.315	0.290	0.311	0.300	0.296	0.291	0.309	0.290	0.105

REFERENCES
Abd.El-Rahman,A.O.; Azab,
A.M.H.; Aggour,A.M.;
Fatma,A.A. Moussa and
Mana Abo.El-Yazid
(2007): "Studies of cellular
and humoral immune
respons in cattle against
FMD bivalent vaccine".
J.Egypt Vet.Med.Assoc., 67
No.2: 265-272.

Abd El-Rahman, A.O.; Faraag, M.A.; Samira El-Kilany; Ali, S.M. and Manal Abo El-Yazied (2006): "Isolation and identification of serotype O of foot and mouth disease virus from imported bulls and its correlation to the current used vaccine strain O1/3/1993". Proc. 3<sup>rd</sup> Inter. Conf. Vet. Res. Div., NRC, Cairo, Egypt, 91-100.

Abeer,E.; Halima,E.;
Eman,M.; Laila ElShehawy and Daoud,A.M.
(2002): "Comparison
between cellular and
humoral immunity of sheep
vaccinated, challenged and
infected by foot and mouth
disease virus". 6<sup>th</sup> Vet. Med
Zag.Conf.(7-9 Sept., 2002),
Hurghada.

Abu Bakr M.A. Aggour (1996): "Studies on FMD vaccination in buffaloes". Ph.D.Thesis (Infectious Diseases), Fac. Vet .Med., Cairo University.

Armstrong, R.M. (1998): detection "The antibodies against foot and mouth disease virus in sheep milk". J.Virological Methods, 169 (1-2): 45-51. Assem Abu Bakre (2006): "Some studies on FMD maternal immunity goats". M.V.Sc Thesis, Fac. Med., Vet. Alexandria University.

Barteling, S.J. and Vreesijk, J. (1991): "Developments in foot and mouth disease vaccines central veterinary institute". Vaccine, 9: 75-88.

Daoud, A.M.;
Abeer, E.Mohamed;
Aggour, A.M.; Waffa El-Said Deghidy and Laila
El-Shehawy (2003):
"Comparative studies on
the immune response of
buffaloes to FMD vaccine
adjuvanted with ISA206 or
aluminum hydroxide gel".
3<sup>rd</sup> Int.Sci. Conf., Mansoura.

El-Nakashly,S.A.; Abou
Zaid,A.A.; Samira El-Kilany
and Abd El-Aty,M.M. (1996):
Isolation and identification of
foot and mouth disease virus
during an outbreak of 1993 in
Egypt". 7<sup>th</sup> Sci.Cong.,
Fac.Vet.Med., Assiut
University, pp. 679-687.

El-Watany, H.; Shawky, M.; Roshdy, O.H. and Samira El-Kilany (1999): "Relationship between cellular and humoral immune response in animals vaccinated with FMD vaccine". Zag. Vet.J., Vol. 27, No.1: 137. Aggour, A.M. Farag, A.M.; and Daoud, A.M. (2005a): "ELISA as a rapid method for correlation the detecting between the field isolates of foot and mouth disease and current used vaccine strain in Egypt". Vet.Med.J.Giza, Vol. 53, No. 4, pp.949-955.

Farag,M.A.; Samira El-Kilany and Abd El-Rahman,A.O. (2006): "The impact of live animal importation on theepizootiology of foot and mouth disease in Egypt". 8<sup>th</sup> Sci.Vet.Med.Conf., Fac.Vet.Med., Zag. University, pp. 351-359.

Farag,M.A.;Shawky,M. and Daoud,A.M.(2005b):

"Western blot incomparison with ELISA for detecting antibodies against foot and mouth disease virus". Vet.Med.J.Giza,Vol.53, No. 4, pp 956-966.

Fathia Abdel-Fattah (2003): "Vaccination of goats with foot and mouth disease vaccines". Master Thesis (Infectious Diseases), Fac. Vet. Med., Alexandria University. Ferria, M.E.V(1976):

"Peudade microneutralization por ae studios de anticuerpos dela fiebre aftosa". Bol. Centro Pan Americano de Fiebre Aftosa (21/22): 17-24.

Kitching, R.P. and Hughes, G.J. (2002): "Clinical variation of foot and mouth disease: sheep and goats". Rev. Sci. Tech. Off. Int. Epiz., 21(3): 505-512.

Moussa, A.A.M.; Ibrahim, M.H.; Daoud, A. and Hussien, K. (1974): "Foot and mouth disease vaccine production in baby hamster kidney (BHK21) cell in suspension in Egypt". Bull.Off.Inst.Epiz., 81 (11-12): 1043-1054.

# El-Sabagh, I.M. et al.

Moussa, H.A.A. and Youssef, N.M.A. (1998): "Serological screening for some viral diseases antibodies in camel sera in Egypt". J.Agric.Res., 76: 867-873. (2006): "Disease OIE information".16 February, 2006. informationdept@oie.net, 19(7): 16-19. Shawky, M.; Ismail, I.; Abdullah, M. and Aggour, A.(2006): "Serosurveillance of FMD in camel, sheep and goat in

area of Halayib and Shalateen".Zag.Vet.J. (ISSN. 110-1458), Vol. 34, No. 2 (2006), pp. 92-97. Starver, P.I.; Classens, A.M.J.M. and Van Bekkum, J.G. (1970): "Some properties of carrier stains of foot and mouth disease virus". Arch Ges Virus Forsch, 29: 113-126. Warnery, U. Kaaden O.R (2004): "Foot and mouth disease in camelids:a review". Vet.J.; 168(2): 134-142.